EFFECT OF COMPACTIN ON THE INCORPORATION OF MEVALONOLACTONE INTO GIBBERELLIC ACID BY GIBBERELLA FUJIKUROI

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Key Word Index—Gibberella fujikuroi; compactin; gibberellin A3; biosynthesis; inhibition.

Abstract—In Gibberella fujikuroi, strain GF-1a, the effect of the sodium salt of compactin on the incorporation of both radiolabelled acetate and mevalonate into gibberellic acid has been investigated. In each case, a concentration of 40 mg/l. caused a significant reduction in the incorporation.

INTRODUCTION

The fungal metabolite compactin (1a) has been isolated from strains of *Penicillium brevicompactum*. It has been shown to be a highly potent, competitive inhibitor of hydroxymethylglutaryl coenzyme-A reductase and a cholesterol-lowering agent in several animals [1]. Thus, formation of cholesterol from acetate is inhibited by compactin in these systems, whereas that from mevalonate is unchanged.

In the course of studying the metabolism of ¹³C-labelled mevalonate in strains of *Gibberella fujikuroi* we wished to maximize the specific incorporation of the precursor into the fermentation products, which include several gibberellins. Hence the effect of compactin on incorporation of mevalonate into gibberellins, particularly GA₃, was of interest to us.

RESULTS AND DISCUSSION

Table 1 shows that, as expected, the incorporation of $[2^{-14}C]$ mevalonolactone into GA_3 was much higher than that of sodium $[2^{-14}C]$ acetate. By comparison of the figures for percentage incorporation into GA_3 in the presence and absence of the sodium salt of compactin (1b)

$$R = \begin{pmatrix} 0 & CH_2 \\ H & H \end{pmatrix}$$

the following results are also apparent. Incorporation of acetate was inhibited by $64.8\,^{\circ}_{\circ}$. However, incorporation of mevalonate was also inhibited, by $38.5\,^{\circ}_{\circ}$. These two figures show that $57.2\,^{\circ}_{\circ}$ of acetate fed was converted to mevalonate, in the presence of, compared with in the absence of, compound 1b.

Table 1. Incorporations of sodium $[2^{.14}C]$ acetate and (3R)- $[2^{.14}C]$ mevalonolactone into GA_3 by Gibberella fujikuroi strain GF-1a, in the presence and absence of 1b

Compounds fed	% incorporation	Average on incorporation
[2-14C]Acetate only	0.66	0.71
	0.76	
	0.70	
[2-14C]Acetate + 1b	0.39	0.25
	0.08	
	0.27	
[2- ¹⁴ C]Mevalonolactone only	12.70	12.35
	12.35	
	12.00	
[2-14C]Mevalonolactone + 1b	7.20	7.60
	7.60	
	8.00	

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Therefore, when Gibberella fujikuroi was incubated with 1b at a concentration of $40 \, \text{mg/l.}$, GA_3 production was inhibited from mevalonate as well as from acetate. This result is in contrast to the analogous situation for the biosynthesis of cholesterol from these precursors which has been reported $\lceil 1 \rceil$ for several animals.

EXPERIMENTAL

Preliminary expts were undertaken using incubations of compactin or its Na salt (1b) (formed by stirring compactin in 0.1 M NaOH for 2 hr at 50°) in 250 ml shake flask cultures of Gibberella fujikuroi, strain GF-1a. Analysis by GC (of Me esters, TMSi ethers) of the metabolites produced after 5 days showed the following. (1) Neither 1a nor 1b affected qualitatively the metabolites formed: the major ones being GA₃, GA₁₃ and fujenal diacid in the ratio 20:6:9. (2) Metabolite levels were decreased by up to 90% by addition of the Na salt 1b. (3) Compound 1b was much more effective in reducing metabolite levels than 1a. (4) The optimum dose of 1b appeared to be ca 40 mg/l.

Consequently, incorporations of sodium $[2^{-14}C]$ acetate and $[2^{-14}C]$ mevalonolactone into GA₃ were measured at 40 mg/l. of 1b, as follows.

Aq. sodium $[2^{-14}C]$ acetate (138 kBq) was added to a 250 ml shake flask containing sterilized 0% ICI medium [2] (50 ml) and 1b (2.0 mg). Freshly Buchner-filtered and H_2O -washed mycelium

from a 4-day-old culture of Gibberella fujikuroi, strain GF-1a, was then added. As a control, the above expt was repeated but without the addition of 1b. The incubations with added $[2^{-14}C]$ mevalonolactone (EtOH soln of 3-(RS) isomers, 62.5 kBq/250 ml shake flask) were performed under identical conditions. All incubations were done in triplicate (total of 12). After shaking for 5 days at 200 orbits/min the mycelia were filtered off and 100 mg cold GA₃ added to each filtrate. The filtrates were then adjusted to pH 2 with 2N HCl, extracted with EtOAc (3 × 50 ml), washed with H_2O (50 ml), concd in vacuo, and chromatographed by prep. TLC (Merck Kieselgel 60 HF₂₅₄; 0.8 mm thick; EtOAc-CHCl₃-HOAc, 15:5:1), eluting the band at R_f 0.45 using H_2O -satd EtOAc. The resultant GA_3 was then crystallized from Me_2CO -petrol to constant specific radioactivity. The results are indicated in Table 1.

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TRITERPENOIDS FROM HYPTIS SUAVEOLENS ROOTS

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Key Word Index—*Hyptis suaveolens*; Labiatae; triterpenoids; α -amyrin; β -amyrin; 3β -hydroxylup-12-en-28-oic acid.

Abstract—A new natural triterpenoid, 3β -hydroxylup-12-en-28-oic acid, has been isolated from the roots of *Hyptis suaveolens* in addition to α - and β -amyrin.

INTRODUCTION

In continuation of our earlier work [1-3] we have isolated three more triterpenoids from the roots of *Hyptis suaveolens*, and their structural elucidation is described in the present communication.

RESULTS AND DISCUSSION

The first compound was shown to be α -amyrin by mp, IR and colour tests [4–7]. This was confirmed by co-TLC with an authentic sample. Similarly the second compound proved to be β -amyrin.

The third compound, mp 288–289° (alcohol), $[\alpha]_D^{30}$

 $+12^{\circ}$ (pyridine), gave colour reactions [4–7] which indicated that it is unsaturated. Elemental analyses and MW (456) determination by mass spectrometry gave the molecular formula $C_{30}\,H_{48}\,O_3$. The IR spectrum showed hydroxyl (3440 cm⁻¹), carboxyl (1690 cm⁻¹), unsaturation (1645 cm⁻¹) and gem-dimethyl (1380 and 1370 cm⁻¹) functions in the molecule. Also, the –COH stretching bands at 1035 and 1025 cm⁻¹ showed that the hydroxyl group is equatorial and is present at the C-3 position of an A/B trans-triterpenoid [8].

In the ¹ H NMR spectrum, singlets at δ 0.66 (6H, s), 0.76 (3H, s), 0.90 (3H, s) and 1.04 (3H, s) demonstrated five tertiary methyl groups [9]. Two doublets appearing at